

**COURSE DATA****Data Subject**

Code	36700
Name	Plant Molecular Biology
Cycle	Grade
ECTS Credits	4.5
Academic year	2021 - 2022

Study (s)

Degree	Center	Acad. year	Period
1109 - Degree in Biochemistry and Biomedical Sciences	Faculty of Biological Sciences	3	Second term

Subject-matter

Degree	Subject-matter	Character
1109 - Degree in Biochemistry and Biomedical Sciences	7 - Biología vegetal	Obligatory

Coordination

Name	Department
MUÑOZ BERTOMEU, JESUS	25 - Plant Biology

SUMMARY

Photosynthetic organisms are at the base of the food chains and are probably the living beings that have contributed most in the past to create the environmental conditions for the development of the rest of aerobic living beings. Hence, a challenge for future generations of molecular biologists is the global understanding of them, in order to develop the biotechnological potential of plants to improve the environmental quality of life on Earth.

Plant Molecular Biology will be studied from the molecular point of view, as this group of organisms, and in particular the higher plants have common characteristics that differentiated form a separate knowledge field and is the subject of this course.

First, the differential characteristics of the plants will be studied. Furthermore, the comparison of sequenced genomes of several higher plants enables functional analysis of plant genomes with respect to other groups of organisms. The plant nuclear genome and its intercommunication with organellar genomes will be also studied, in addition to the genetic and molecular tools that allow progress in the current study of plants and their application in biotechnology with biomedical purposes.



The basic mechanisms that distinguish plants from other organisms, such as water acquisition and mineral elements uptake, photosynthetic CO₂ fixation and its distribution throughout the plant. Finally, the molecular mechanisms and gene expression changes that regulate the processes of development. Furthermore, the knowledge of the molecular bases of plants and their genetic transformation methods, allows the design of biotechnological strategies aimed at different purposes of interest, due to the economic and environmental importance of plants.

Plant Molecular Biology is a subject with an important experimental content, so four laboratory sessions, mainly aimed to familiarize students with the recombinant DNA technology applied to plants and their genetic transformation, mediated by *Agrobacterium tumefaciens*, will be studied.

PREVIOUS KNOWLEDGE

Relationship to other subjects of the same degree

There are no specified enrollment restrictions with other subjects of the curriculum.

Other requirements

There are no specified restrictions on enrollment with other subjects of the curriculum.

OUTCOMES

1109 - Degree in Biochemistry and Biomedical Sciences

- Have capacity for analysis, synthesis and critical reasoning in the application of the scientific method.
- Have an integrated view of normal and altered cell function, including metabolism and gene expression.
- Students must have acquired knowledge and understanding in a specific field of study, on the basis of general secondary education and at a level that includes mainly knowledge drawn from advanced textbooks, but also some cutting-edge knowledge in their field of study.
- Students must be able to apply their knowledge to their work or vocation in a professional manner and have acquired the competences required for the preparation and defence of arguments and for problem solving in their field of study.
- Students must have the ability to gather and interpret relevant data (usually in their field of study) to make judgements that take relevant social, scientific or ethical issues into consideration.
- Students must be able to communicate information, ideas, problems and solutions to both expert and lay audiences.
- Students must have developed the learning skills needed to undertake further study with a high degree of autonomy.



- Be able to assimilate scientific texts in English.

LEARNING OUTCOMES

SKILLS TO ACQUIRE:

- Ability in handling basic literature sources related to the subject and skill to deepen their knowledge on a specific topic related to plants, for critical analysis of scientific articles.
- To understand and use the proper terminology for describing molecular processes in plants.
- Skill in handling molecular and bioinformatics tools aimed to know the functions of plants and to facilitate their applications in biomedicine.
- Skill in handling and identification of lines with gain and loss of function in relation to experiments that arise in practical classes.

SOCIAL SKILLS:

- To develop the scientific language needed to communicate the basics of the subject and ability to argue with rational criteria in the context of a scientific discussion related to the subject.
- Ability to teamwork and use of new communication technologies.
- Capabilities with updating of the existing information on a specific problem, adequate temporary organization, order and analyze such information critically and publicly expose and defend the work.
- Ability to experimental work, appropriate interaction with peers and teacher laboratory and development of the critical capacity of the experimental results.

DESCRIPTION OF CONTENTS

1. Introduction

Why study molecular biology of plants in the Biochemistry and Biomedicine degree?. Differential molecular characteristics of plants.

2. The nuclear genome

Characteristics of the plant nuclear genome: ploidies and mobile genetic elements. Plant genome sequencing: functional annotation. Regulation of plant nuclear genomes: RNA polymerases and their regulation; epigenetic mechanisms of gene regulation. Small RNAs regulators



3. Cytoplasmic genomes

Features, organization and functionality of the plastome and chondriome. RNA polymerases and promoters of plastid genes. Anterograde and retrograde communication between genomes.

4. Tools in molecular biology and genetics of plants

Model photosynthetic organisms. Genetic and electronic resources. Nuclear and chloroplast transgenesis. Collection, and analysis of mutant types. Omics technologies in plants. Biotechnological applications related to biomedicine.

5. Movement of water and solutes in plants

Transpiration and regulation. Absorption of water and nutrients through the roots and xylem transport. Assimilate transport.

6. Plants as source of energy

Energy absorption by plants: photosystems. Calvin cycle and photorespiration. C4 and CAM metabolism. Sulfur and nitrogen assimilation.

7. Molecular basis of plant hormone action

Main hormonal groups: Auxins, Gibberellins, Cytokinins, Ethylene, Absciscic acid, Brassinosteroids, Estrigolactones, Jasmonates, Salicylates, Polyamines, oligopeptides and Nitric oxide. Signaling by: ubiquitination, kinase type receptors and hybrid kinase systems. Interactions and integration of hormonal signaling pathways.

8. Molecular bases of plant growth and development

Establishment of the apical-basal axis. Structure and maintenance of stem and root apical meristems.

9. Light and temperature as signals of the development of plants

Scotomorphogenesis and Photomorphogenesis. Signaling pathways from the main photoreceptors (Phytochromes, Cryptochromes, Phototropines, Family ZEITLUPE and UVR8). Thermomorphogenesis. The circadian clock.

**10. Molecular bases of flowering**

Flower development: from Model ABC to model ABC (D) E. Photoperiodic control of flowering. Vernalization. Other flowering pathways.

11. Molecular bases of seed development

Pollination and double fertilization. Embryogenesis and development of the endosperm. Maturation of seeds. Dormition.

12. Molecular basis of Senescence.

Monocarpic and Polycarpic Senescence. Abscission of organs

13. Laboratory classes

Sesión 1.- Extracción de DNA genómico de plantas control y mutantes.

Sesión 2.- Identificación de mutantes de inserción de T-DNA y TILLING mediante genotipado por PCR y análisis de restricción.

Sesión 3.- Análisis de la expresión génica en plantas con promotores constitutivos e inducibles mediante el ensayo histoquímico de la -glucuronidasa.

Sesión 4.- Identificación de plantas transgénicas de Arabidopsis de inserción única y homocigotas para el transgen mediante estudio de la segregación de genes marcadores.

WORKLOAD

ACTIVITY	Hours	% To be attended
Theory classes	31,00	100
Laboratory practices	12,00	100
Tutorials	2,00	100
Development of group work	15,00	0
Preparation of evaluation activities	42,50	0
Preparation of practical classes and problem	10,00	0
TOTAL	112,50	

TEACHING METHODOLOGY

The development of the course is divided into:

Theory class: 27 hour sessions where the master class will be mainly used for teaching. The teaching material (pdf powerpoint presentations) used by the teacher as well as the specific bibliography used in each issue, will be accessible in advance in the Virtual Classroom (platform support teaching). The Virtual Classroom is considered the official bulletin of the subject for examination sessions, qualifications



and reporting schedules of exam review. The student must be aware of such communications.

Practical laboratory sessions: they are taught during a week in 4 sessions of 3 hours each. Students will have a booklet or script of practices, facilitated by the teacher in advance through the Virtual Classroom. Students must have read the script before performing practical laboratory sessions. Attendance at practical sessions is mandatory. During the session, the teacher will guide the conducting experiments and assist in the discussion of the results obtained.

Seminars, conferences and other activities: activities that allow students to expand their knowledge on the subject and relate with other disciplines and promote the acquisition other than those acquired in the theoretical and practical skills classes will be developed. One of these activities will be critical analysis of scientific papers, selected by the teachers of the course. This activity aims to student training in reading scientific papers (which necessarily involves reading technical English), bringing him/her closer to the original literature from which new knowledge that enable the development and advancement of biomedical sciences are obtained. This activity, mandatory, will be organized jointly with the other courses in the third year. The preparation, presentation and discussion of articles will take place in groups of 2 students and will be supervised by the teacher through tutorials.

EVALUATION

- Theoretical contents will be evaluated with a written exam consisting of questions related to the program content. The result of this evaluation represent 80% of the final grade for the course. There will be the possibility of taking a midterm exam. For those students that pass the midterm exam, the final exam will only include contents from the second half of the subject (those not included in the midterm exam).
- The contents of the laboratory practices will be assessed through a written exam questions that will score 15% of the final grade. The issues may be related to both the experimental part developed, as with the techniques and methodology discussed during the laboratory sessions. Attendance is mandatory and, if approved, the exam is saved for the second call within the same academic year.
- A minimum score of 4 out of 10 in theoretical and practical exams will be required to pass the course.
- To evaluate the activity of critical analysis of scientific papers the following evaluation criteria will be taken into account. You can also valued integration with other theoretical and practical contents of this or other subjects of the degree. We could get a maximum score of 10 points, 5 points still needed to overcome this activity. The mark obtained will represent 5% of the final grade for each course subjects third participants in this activity. If the student does not reach the minimum score required, suspend the course in which performs this activity. Likewise, the participation of other students in the presentation and discussion sessions, may be considered by the teacher to modulate the final grade for the course.
- Repeaters students who have completed practical classes, but have not passed the course may not attend laboratory classes, although they must retake the exam practices and other assessment activities of the subject.
- The student's active participation in the theoretical and practical classes will be assessed, adding up to 0.5 points to the final grade.



REFERENCES

Basic

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 - Hirt H y Shinozaki K (2004) Plant responses to abiotic stress. Springer-Verlag Berlin Heidelberg. Alemania.
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 - Slater A, Scott N y Fowler M (2003) Plant Biotechnology: The genetic manipulation of plants. Oxford University Press, Oxford.
 - Smith AM, Coupland G, Dolan L, Harberd N, Jones J, Martin C, Sablowski R y Amey A (2009) Plant biology. Garland Science New York. EEUU.
 - Taiz L, Zeiger E (2006). Plant Physiology. 4th. Cumming Publ. Company, Inc.
 - Westhoff, P. et al., (1998). Molecular plant development from gene to plant. Oxford University Press.
 - Yoshioka K y Shinozaki K (2009) Signal crosstalk in plant stress responses. Wiley-Blackwell. New Delhi, India.
- Se usarán revisiones en publicaciones periódicas como el Trends in Plant Science, Current Opinion in Plant Biology y similares, para aspectos concretos del temario.

ADDENDUM COVID-19



This addendum will only be activated if the health situation requires so and with the prior agreement of the Governing Council

1 y 2) Contenidos y Volumen de trabajo.

Se mantienen los contenidos inicialmente recogidos en la guía docente. En cuanto al volumen de trabajo se mantendrá el especificado en la guía docente, pero la presencialidad en caso de confinamiento será menor en función de la duración del confinamiento.

3) Metodología.

El punto de inicio dado el número de estudiantes y las aulas disponibles es de presencialidad del 50% en clases de teoría con retransmisión síncrona, y plena presencialidad en el resto de las actividades. Sin embargo, ante la posibilidad de que la evolución de la situación derivada de la COVID-19 obligue a una reducción de la presencialidad, se tomarán las siguientes medidas:

1) Las actividades presenciales en aula se sustituirían en función de las herramientas tecnológicas disponibles en el aula en el momento de desarrollo del curso, por algunas de las siguientes metodologías:

- Videoconferencia síncrona
- Presentaciones Powerpoint locutadas en Aula Virtual
- Presentaciones Powerpoint con apuntes extendidos en Aula Virtual
- Propuestas de actividades de resolución de Cuestionarios de Aula Virtual y entrega de tareas y cuestiones por Aula Virtual

2) Las actividades presenciales de prácticas de laboratorio, se sustituirían por algunas de las siguientes metodologías:

- Prácticas de laboratorio simuladas mediante videoconferencia
- Presentaciones Powerpoint locutadas en Aula Virtual
- Trabajo con datos experimentales suministrados
- Videos de las prácticas que no se puedan realizar en el laboratorio

3) Para tutorías y dudas se utilizarían las siguientes metodologías:

- Videoconferencias en Aula Virtual



- Comunicación directa profesor-estudiante a través del correo institucional
- Tareas a realizar mediante actividades en el aula virtual

4) Evaluación.

En caso de que los exámenes no pudieran ser presenciales, se realizarían 'on line' en Aula Virtual mediante las herramientas disponibles.

Los detalles concretos de la adaptación a las situaciones que se pudieran producir se supervisarán por la CAT y se comunicaran a los estudiantes a través de Aula Virtual