



## COURSE DATA

Data Subject	
<b>Code</b>	33135
<b>Name</b>	Macromolecular biosynthesis and its regulation
<b>Cycle</b>	Grade
<b>ECTS Credits</b>	9.0
<b>Academic year</b>	2019 - 2020

## Study (s)

Degree	Center	Acad. Period year
1109 - Degree in Biochemistry and Biomedical Sciences	Faculty of Biological Sciences	2 Annual

## Subject-matter

Degree	Subject-matter	Character
1109 - Degree in Biochemistry and Biomedical Sciences	9 - Genética y biología molecular	Obligatory

## Coordination

Name	Department
PEREZ ORTIN, JOSE ENRIQUE	30 - Biochemistry and Molecular Biology
TORDERA DONDERIS, VICENTE	30 - Biochemistry and Molecular Biology

## SUMMARY

"Biosynthesis of macromolecules and its regulation" is a compulsory annual subject of the second year of the degree in Biochemistry and Biomedical Sciences and corresponds to 9 ECTS credits. It is intended to provide students with knowledge about the molecular mechanisms that enable the perpetuation and transmission of genetic information. This subject is part of the molecular biology core, which has undergone a huge development in recent years. His field of study is at the crossroads of Biochemistry, Genetics, and Cell Biology. Its study is essential to understand the processes taking place in the cell at the molecular level.



## PREVIOUS KNOWLEDGE

### Relationship to other subjects of the same degree

There are no specified enrollment restrictions with other subjects of the curriculum.

### Other requirements

## OUTCOMES

### 1101 - Degree in Biochemistry and Biomedical Sciences

- Desarrollo de la capacidad de razonar y aplicar el método científico.
- Comprensión de la lógica molecular de los seres vivos como producto de la evolución.
- Capacidad para trabajar en el laboratorio de genética y biología molecular incluyendo seguridad, manipulación, eliminación de residuos y registro anotado de actividades.
- Conocer y comprender las bases moleculares de la información genética y los mecanismos de su transmisión y variación.
- Relacionar las características estructurales y funcionales de las macromoléculas.
- Conocer los elementos comunes y los diversos de la genética y la biología molecular de los diferentes tipos de organismos vivos.

## LEARNING OUTCOMES

- Acquiring knowledge and understanding of genetics and molecular biology.
- Solving theoretical and practical exercises.
- Practical activities in the laboratory and analysis of the results.
- Generating a written work and oral presentation with visual support.

## DESCRIPTION OF CONTENTS

### 1. Theme 1. Introduction

Stages in the history and development of molecular biology and the central dogma of molecular biology as a science. Definition and field of study of molecular biology. Working methods. Introduction to sources of information. (2 hours)

**2. Theme 2: General features of the replication process**

Semi-conservative nature. Orderly and sequential replication: the origin of replication. Unidirectional or bidirectional replication. (3 hours)

**3. Theme 3. DNA polymerases**

Historical introduction: discovery of DNA pol I of *E. coli*. Catalyzed reactions. DNA polymerase activity: characteristics. Activities 3' exonuclease 5' and 5' 3'. Three-dimensional structure of pol I. Other polymerases of *E. coli*. Features compared with pol I. Complexity of DNA pol III holoenzyme system. Other DNA polymerases. (3 hours)

**4. Theme 4: Semi-discontinuous replication: elements involved**

Okazaki experiences. Characteristics of DNA ligases of *E. coli* and phage T4. Reaction mechanisms. Roles of DNA ligases in vivo. Start of the replicative synthesis in the fork: the RNA as a primer. The primase. The primosome. SSB proteins. DNA helicases. Role of DNA topoisomerases during replication. (4 hours)

**5. Theme 5: The replication complex**

General outline of all components of the replication complex in the replicative fork. The replisome and simultaneous replication of both strands: topological considerations. Initiation of replication in *E. coli* proteins involved. Ori sequences. End of the circular DNA replication. End of non-circular DNA replication. DNA methylation in prokaryotes: restriction-modification systems. (5 hours)

**6. Theme 6: Specific characteristics of the replication in eukaryotes**

Enzymology of replication in eukaryotes. The eukaryotic replication fork: a comparison with prokaryotes. Replicons and replication origins. Controlling initiation of replication. Replication of telomeres: telomerase. Chromatin structure and replication. Replication of DNA in organelles. (3 hours)

**7. Theme 7: Transcription: definition and historical context of its discovery**

Hypothesis of "bridging molecule". Demonstration of the existence of the mRNA. Features mRNA: differences between prokaryotes and eukaryotes. (2 hours)

**8. Theme 8: Transcription in prokaryotes**

The DNA-dependent RNA polymerase: structure and function of the subunits. The prokaryotic promoter. Stages of transcription. Start: sigma factor cycle. Elongation: transcription bubble. Movement of RNA polymerase. Transcription and supercoiling. Chain completion: intrinsic terminators and Rho-dependent protein. (4 hours)



#### **9. Theme 9: Transcription regulation in prokaryotes**

General scheme of the levels of regulation. Regulation of promoters by the sigma factor. Promoters regulated by protein CAP. Cis/trans regulation in prokaryotes. Regulation by positive and negative control, induction and repression. The lactose operon. Regulation of transcription by anti-termination. Regulons. (3 hours)

#### **10. Theme 10: Transcription in eukaryotes**

Differences with transcription in prokaryotes. Eukaryotic RNA polymerases. Chromatin and transcription. Elements necessary for the formation of the preinitiation complex: basal transcription factors and promoters. Transcription initiation by RNA pol I, pol III by RNA and the RNA pol II. Elongation and termination of transcription. (6.5 hours)

#### **11. Theme 11: Regulation of transcription in eukaryotes**

Differences at the molecular level between prokaryotes and eukaryotes. Levels of regulation. Concept of active and inactive chromatin. Regulatory mechanisms associated with the chromatin structure. DNA methylation as a regulator. Cis/trans regulation in eukaryotes: some cases. Nuclear organization of transcription. Transcriptional eukaryotic genome organization: redefining the gene. (5.5 hours)

#### **12. Theme 12: Post-transcriptional modification of the RNA**

Types of RNA processing. Precursors of the rRNAs and tRNAs. Role of snoRNAs. Correction of the mRNA sequence. Processing of mRNA in eukaryotes. Splicing. Autocatalytic introns. Ribozymes. (4.5 hours)

#### **13. Theme 13: Messenger transport and stability**

Transport of mRNA to the cytoplasm. mRNA stability. Degradation of defective mRNAs in the nucleus and cytoplasm. Role of the poly-A in regulation. (2.5 hours)

#### **14. Theme 14: The genetic code and the translation machinery**

Features. Exceptions to the universal code. Suppressor tRNAs. Selective use of codons. Origin and evolution. Current components. The tRNA: pairing rules. The ribosome: structure and components. Aminoacyl-tRNA sintetasas. Evolutionary origin of the translation process.(3 hours)

**15. Theme 15: Steps in the translation process**

Initiation in prokaryotes. Initiation in eukaryotes. Elongation. Energy cost of translation. Termination. Translation-inhibiting antibiotics. Energy balance of translation. Post-translational modifications(2.5 hours)

**16. Theme 16: Regulation of translation in prokaryotes**

Autogenous regulation of the synthesis of ribosomes. Structures in the mRNA leader: riboswitches and attenuation. Restrictive response. Ribosomal slips and jumps. (1.5 hours)

**17. Theme 17: Regulation of translation in eukaryotes**

Regulation of initiation: eIF4G, eIF4E, eIF2. Pathways of response to external stimuli. Restrictive response for GCN4. Subcellular localization of mRNA. (1.5 hours)

**18. Theme 18: Regulation by RNA**

Antisense RNA as a regulatory mechanism in prokaryotes. Riboswitches. Antisense RNA in eukaryotes: RNA interference and post-transcriptional silencing. Cryptic transcription. Long non coding RNA lncRNA.(2 hours)

**19. Theme 19: Integration of mechanisms for gene expression regulation**

Role of regulation in each of the stages. Evolutionary and functional reasons for the existence of multiple stages of regulation. Some examples of multistage regulation: the case of the lambda phage, the change of sex in yeast and examples of control of embryonic development in animals. (1 hour)

**20. Theme 20: Quality control of genetic information**

Problem overview. Expression strategies: quality, cost and noise. Fidelity polymerases: mechanism of correction. Accuracy of translation: mechanisms of control during the stages. Structural information. Response mechanisms to alterations in protein sequence. Response mechanisms to changes in protein structure. (1 hour)

**21. Laboratory experiences**

PRACTICE 1. Chromatin structure.

PRACTICE 2. Checking for the presence of an intron in the gene ACT1 of the yeast *Saccharomyces cerevisiae*.

PRACTICE 3. Study of the regulation of the synthesis of -galactosidase enzyme in *Escherichia coli*.

PRACTICE 4. Subcellular localization and characterization of the high affinity transporter glucose Snf3p.



## WORKLOAD

ACTIVITY	Hours	% To be attended
Theory classes	68,00	100
Laboratory practices	16,00	100
Classroom practices	6,00	100
Development of group work	20,00	0
Study and independent work	55,00	0
Readings supplementary material	7,00	0
Preparing lectures	30,00	0
Preparation of practical classes and problem	20,00	0
<b>TOTAL</b>	<b>222,00</b>	

## TEACHING METHODOLOGY

The development of the subject is divided into:

**Theoretical classes:** A total of 60 sessions of one hour each are needed to cover the program. The teacher will present the relevant contents of the course, using audiovisual equipment for fast and consistent acquisition of the main concepts. The material will be available in advance in the institutional web page.

**Problem sessions:** 6 one-hour sessions will be held throughout the course, usually at the end of each of the sections of the agenda. These sessions reinforce the concepts presented in the theoretical sessions and stimulate the active participation of students through the resolution and group discussion of specific issues. The teacher will prepare a series of questions for each topic or subject block that students will work individually (at home) and collectively (through exposure and discussion of them in group class). The teacher may request the delivery of some of the problems, prior to the sessions. In these cases, the work will be submitted electronically through Virtual Classroom. For discussion of the problems, students will be notified in advance of the date and the issues that will be brought about for discussion.

**Hands-on labs:** They are of compulsory attendance. The delivery of a written description is also mandatory. They will be done in 4 lab sessions, adding up to 16 hours.

**Seminars:** This activity will be organized jointly with the other subjects of the second degree course. The activity is the preparation and presentation for approximately 30 minutes by the students (in groups of two students) of a specific topic related to the subject. Students will complete the preparation and presentation of the seminar once during the school calendar. Their active participation in the discussion of other seminars can be taken into account. In "Biosynthesis of macromolecules and its regulation" 7 workshops, whose themes will be proposed each course by the teachers, will be included. Also a seminar of a visiting scholar will be held during lecture hours. The workshop activities will be compulsory.



## EVALUATION

There will be two qualifying exams of the theoretical material. The first will assess acquisition of the contents for the first half of the agenda and will take place halfway through the academic year. The second test at the end of the lecturing period will cover the second half of the theoretical contents agenda. Each test is scored on 4.25 points maximum. The exam of a party can be kept until the second call if the rating is greater than 1.5.

The total final grade comprises 85% of the test scores for theory (8.5 points), 10% of the score from lab work evaluation (1 point) and 5% of the mark of the seminar presented in any of the second-year subjects (0.5 points). The rating in practices (1 point) will result from the grade earned during the work at the lab: written memory and specific test questions included in the second exam of the subject. The mark to be obtained in the seminar will be derived from the capacity of synthesis and integration of information shown by the student, the clarity and quality of the oral presentation and the debate generated around the questions raised by students and teachers. The scores for the lab written report and the seminary (up 0.5 points) will be saved for the second call.

To pass the course it will be essential to have made the practice (except those students who have performed in previous years, in which case it is not necessary to repeat the practices but the specific test questions) and participate in seminar activities. To pass the course will need to obtain a final score less than 5 out of 10 and have passed the grade of 1.5 in each of the two tests of theory.

## REFERENCES

### Basic

- CLARK, D.P. (2010). Molecular Biology: academic cell update. Elsevier
- LEWIN, B. (2010). "Genes X". Jones & Bartlett.
- TORDERA, V., DEL OLMO, M., MATALLANA, E., PÉREZ ORTÍN, J.E. (2007). Qüestions en Biologia Molecular. Collecció Educació Laboratori de Materials. Universitat de València.
- TROPP, B. E. (2008). Molecular Biology: Genes to Proteins. 3<sup>a</sup> ed. Jones & Bartlett.
- WATSON, J.D., BAKER, T.A., BELL, S.P., GANN, A., LEVINE, M. y LOSICK, R. (2008). Molecular Biology of the Gene (6<sup>a</sup> ed.). Pearson/Benjamin Cummings.
- WEAVER, R.F. (2008). Molecular Biology (4<sup>a</sup> ed.). McGraw-Hill International.

### Additional

- ADAMS, R.P.L. "DNA Replication". (1991) IRL Press (série "In focus").
- ARNSTEIN, H.R.V. y COX, R.A. (1992). Protein Biosynthesis. IRL Press. (serie "In focus").
- BEEBEE, T., BURKE, J. (1992). "Gene structure and transcription". 2a ed. IRL Press (serie "In focus").
- COX, T.M., SINCLAIR, J. (1997). Molecular Biology in Medicine. Blackwell Sciences, Oxford.
- DARNELL, J. E. (2011). RNA: life's indispensable molecule. Cold Spring Harbor Laboratory Press.
- ELLIOTT, D. LADOMERY, M. (2011). Molecular Biology of RNA. Oxford University Press.



- KORNBERG, A., BAKER, T.A. (1992). "DNA replication". 2a ed. Freeman.
- LATCHMAN, D.S. (1991). "Eukaryotic Transcription Factors". Academic Press.
- LEON -SERRANO, J.L. y GARCIA-LOBO, J.M. (1990). "Manual de Genética Molecular". Ed. Síntesis.
- LODISH, H., BALTIMORE, D., BERK, A., ZIPURSKY, S.L., MATSUDAIRA, P., DARNELL, J. (1995). "Molecular Cell Biology". 3<sup>a</sup> ed. Scientific American Books.
- LUQUE, J., HERRAEZ, A. (2001) Biología Molecular e Ingeniería Genética. Conceptos, técnicas y aplicaciones en Ciencias de la Salud. Ediciones Harcourt S.A.
- SINGER, M., BERG, P. (1991) Genes and genomes. University Science Books, Mil Walley, California
- STENT, G.S. y CALENDAR, R. (1981). "Genética Molecular. Una introducción narrativa". Ed. Omega.
- VALPUESTA, J.M. (2011). A la búsqueda del secreto de la vida. Una Breve Historia de la Biología Molecular. Editorial Hélice-CSIC.

## ADDENDUM COVID-19

**This addendum will only be activated if the health situation requires so and with the prior agreement of the Governing Council**

### 1. Contenidos

Se mantienen los contenidos inicialmente recogidos en la guía docente

### 2. Volumen de trabajo

El volumen de trabajo se mantiene

Se mantiene la programación de las clases de teoría en las fechas y horas previstas y su duració

Las clases de prácticas, a partir del 15 de marzo pasan a ser no presenciales por lo que no se mantienen los horarios previstos. Sin embargo, tal y como se explica en el apartado siguiente (Metodología Docente) las clases prácticas presenciales son sustituidas por un trabajo semejante, con objetivos semejantes y con una duración aproximada equivalente al de las prácticas presenciales, solo que hecho de forma no presencial.

### 3. Metodología docente

Se pasa al formato on-line a partir del día 15 de marzo hasta el final del curso. Desde esa fecha:

-Se usa el formato de diapositivas con explicación grabada para las clases de teoría y cuestiones. Se suben copias en PDF de las presentaciones a Aula Virtual así como la resolución de las cuestiones propuestas. Las consultas se resuelven en un foro de Aula Virtual.

-Las clases de prácticas presenciales se sustituyen por un trabajo no presencial que plantea los mismos experimentos que las prácticas presenciales. Se trata de proporcionar al estudiante un protocolo detallado de cada experimento, así como unos resultados brutos en forma de tablas y figuras a partir de los cuales



los estudiantes deben elaborar un trabajo con los resultados presentados en forma de figuras y respondiendo a una serie de cuestiones.

Para ello se ha subido al Aula Virtual una serie de materiales complementarios con explicaciones. Además, se crea un Foro en Aula Virtual “Prácticas no presenciales” para resolver las dudas específicas de esta actividad

#### 4. Evaluación

Dado lo extraordinario de la situación y la generalización de los exámenes online, apelamos a la responsabilidad y a la ética de los estudiantes durante su realización. Si se detectara algún intento de copia u otro tipo de fraude, se adoptarán con rigor las medidas disciplinarias aplicables en estos casos

-Se ha incrementado el peso de los Seminarios (por ser estos una actividad desarrollada de forma continuada a lo largo del Curso), pasando de 0,5 puntos a 1 punto.

-Se ha incrementado el peso de las prácticas pasando de 1 punto a 2 puntos distribuidos de la siguiente forma: 1 punto de la elaboración de una Memoria de resultados de las prácticas, ya sea en forma presencial (un grupo de prácticas pudo hacerlo antes de la pandemia) como no presencial, y 1 punto de la contestación a una prueba objetiva (un examen de tipo test) hecho en Aula Virtual, sobre el material suministrado como Prácticas no presenciales.

-Los dos parciales de la asignatura pasan a tener un valor cada uno de ellos de 3,5 puntos, siendo realizado el segundo de ellos mediante un examen de tipo test en Aula Virtual. La nota del examen de una de las partes se podrá conservar hasta la segunda convocatoria si la calificación es superior a 1,2 (sobre 3,5).

-La parte correspondiente a las teorías, cuestiones y examen de prácticas de la segunda convocatoria se realizarán mediante exámenes de tipo test en Aula Virtual.

-Si por causas técnicas algún estudiante no pudiera realizar el examen online, se realizará una prueba alternativa de tipo ORAL

-Con el fin de garantizar que las calificaciones de este curso no serán globalmente diferentes de las de cursos presenciales se ajustará la media global de las calificaciones a la media global de las calificaciones de los últimos 3 cursos.

#### 5. Bibliografía

La bibliografía recomendada se mantiene